Confocal Quantitative Image Cytometer



Specifications

Item	Specification
Optics	Microlens enhanced dual wide Nipkow disk confocal
Fluorescence	Laser : Choose Max.4 lasers from 405 / 488 / 561 / 640 nm
	EM Filter : Max. 10 filters
Transmitted illumination (Option)	Phase contrast, Bright field Light source : LED
Camera	sCMOS 2560×2160 pixel, 16.6×14.0 mm
Objective lens	Max.6 lenses
	Dry : 2x, 4x, 10x, 20x, 40x Long working distance : 20x, 40x Phase contrast*1 : 10x, 20x
Attachment	All wells imaging type, Chambered type* ²
Sample vessel	Microplate (6, 12, 24, 48* ³ , 96* ³ , 384* ³ , 1536* ³ well), Slideglass* ^{4*5} , Cover glass chamber* ⁴ , Dish* ⁴ (35, 60 mm)
XY stage	High-precision XY stage, designated resolution: 0.1 µm
Stage heater (Option)	Stage heater with chamber
	Controllable temperature range : Room temperature $+5 - +17$ °C, Max.40 °C
	Settable temperature resolution: 0.1 °C
7 feeue	Fundicity keeping time: Over 6 hours
2 TOCUS	Electric 2 motor, designated resolution: 0.1 pm
Fosturo data	Laser autorocus, sontware autorocus
Data format	Cantinger of the state of the s
Data Iomat	Output image from to TIFE (16 bit A bit) PNG IPEG
	Output movie format · WMV_MP4
	Output numerical data format : ECS. CSV. ICE
Workstation	Measurement and analysis workstation
Gas Mixer (Option)	CO2 concentration : Atmospheric concentration – 7 %
	O2 concentration : 3 % – Atmospheric concentration
Size/Weight	Main unit : $600 \times 400 \times 298$ mm, 43 kg (Standard model) , $600 \times 400 \times 437$ mm (With Phase contrast option)
-	Utility box : $275 \times 432 \times 298$ mm, 18 kg
	Gas Mixer (Option) : 170 × 260 × 280 mm, 5.2 kg
Environment	Main unit and Utility box : 15 – 35 $^{\circ}$ C, 20 – 70 % RH No condensation
	Gas Mixer (Option) : 20 – 30 $^{\circ}$ C, 10 – 85 % RH No condensation
Power consumption	Main unit and Utility box : 100-240 VAC, 800 VAmax
	Workstation : 100-240 VAC, 650 VAmax
	Gas Mixer (Option) : 100-240 VAC, 40 VAmax
*1 Phase contrast option is required *2 *4 Sample holder option is required *5	Stage heater option is required to use environment keeping function *3 Phase contrast observation is unavailable Environment keeping function is unavailable



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Cell measurement by high-throughput 3D imaging

Confocal Quantitative Image Cytometer **CQ**¹ offers a new approach to cell measurement



Clear 3D images obtained from confocal microscopes have been enabling advancements in cell biology research for many years. This imaging technology combined with population analysis now provides a significant advancement for cytometry.

The CQ1 enables clear 3D imaging, object recognition, and rapid quantification of live cells and cell clusters. Images linked to data help in the understanding, and enhance the reliability of data. The CQ1's live cell chamber acts as a cellular incubator enabling time lapse imaging while the CQ1's unique imaging technology is gentle on the cells.

The Yokogawa CQ1 is an easy to use all-in-one confocal microscope for a reasonable price. The CQ1 comes with a number of configurable options and can be integrated into a fully automated screening system.







Measurement

Recognition

Ouantification



Enables measurement of spheroids,

- Possible to measure cells in culture dish without preprocessing such as cell peeling, unlike a flow cytometer
- Thanks to the confocal disk confocal, 3D images are acquired rapidly and gently
- Max.10 colors emission with 4 colors excitation and transmission illuminataion imaging
- Live cell chamber and time-lapse measurements
- Rich feature extraction to facilitate sophisticated cell analysis
- Wide field of view and tiling capability enables easy imaging of large sample

Offers the similar capabilities as flow cytometery

- Analyzed data displayed in real-time with image acquisition (On the fly analysis)
- Application protocols guided by templates
- Ability to trace back to the original image from a data point in a graph and to remeasure
- All-in-one system with easy operation

Open platform

- Expandable to integrated systems as an image acquisition and quantification instrument
- Output FCS/CSV/ICE data format readable by third-party data analysis software
- Connectable with external systems via plate handling robot
- A variety of cell culture and sample vessels are applicable

Compact footprint, light weight bench-top device; no need for darkroom

■ Contrast of measurement methods

Flow cytometer

Non-confocal imaging system

- Cell peeling treatment is necessary. Risk of damaging to cell







Unable to re-measure nor

confirm by image

Imaging is difficult sample is thick.



colonies and tissue sections.





Confocal imaging system





3D imaging of thick sample

In addition, CQ1 is high-throughput and gentle with cell.

Multiple Functions Fully Integrated in a Compact Box

■ Confocal Scanner Unit

Multi-beam scan by "Microlens enhanced dual Nipkow disk confocal" achieve high-throughput 2D/ 3D imaging with minimum damage to samples.



Example: Cellcycle measurement of cancer cells

Microscope Unit

Maximal performance objective lens (super apochromat) and the widest field/ highest-resolution sCMOS camera achieve high-throughput measurement of submicron sample.

■ Emission Filter

Up to 10 Emission filters can be mounted. Measurement of multiple markers can be achieved in just one experiment.

■ Illumination for Fluorescent Imaging (Laser)

can be mounted.





Environment Keeping Function

Stage heater controls temperature and humidity and gas mixer controls CO₂ / O₂ concentration of sample environment.

Measurement with keeping cell viability can be achieved by this function.



Supported Sample Vessels Various sample vessels, as microplate, can be used for measurement. Microplate 35 mm dish^{*1} 60 mm dish*1

Slide glass*1

Cover glass chamber*1







*1 Option *2 Under development *3 GM-8000 (Tokai Hit) is recommended *4 Output by OME-TIFF format *5 Output by FCS, CSV or ICE format *6 Example of system integration. CQ1 also has image / numerical analysis function. *7 StrataQuest is the trademark of TissueGnostics *8 FCS Express[™] 5 Image Cytometryis is the trademark of Denovo Software



Up to 4 laser illumination for confocal (fluorescent) imaging

Measurement of multiple fluorophore can be achieved in just one experiment.





■ Illumination for Transmission Imaging^{*1} Transmission illumination for phase contrast or bright field*2 imaging can be mount. It is useful for confirming sample shape.

Set the Condition and One Click!

-Easy & Universal Software-









Let's start the easiest 3D Measurement!

The CQ1 is the easiest way 3D measurement system Simple cell identification, colony counting, and complex colony property analysis are available. Of course you can do whole well imaging and analysis.



Quality control: Induction of differentiation High-speed 3D resolution Multi-color



Aggregated cell images were taken in slices and presented as 3D.

Marker expression level as well as spatial information of individual cells were quantified via image analysis.



Template Spheroid structure Cell-by-cell measurement of aggregated cells like spheroids.

Applications Sheroids, Differentiation



■ Example of protocol

Image data

(Whole well, 3D)











Differentiation marker positive





Quality control: human iPSC sphere

Cell-by-cell measurement of aggregated cells like spheroids

Want to go more deeper analysis!

High-quality Confocal images from the CQ1 enable many types of image analysis. Morphology change, particle analysis and other HCA that require high resolution images.

Of course CQ1 can work like simple Confocal Microscopy to get analyzed data and images.





Analysis: gamma-H2AX focus formation





The phosphorylation of histone H2AX Ser139 (gamma-H2AX) is one of the significant events upon the breakage of double strand DNA. Quantitative measurement of gamma-H2AX focus formation can be easily performed by using the high-speed confocal image acquisition in combination with the Granule Analysis Template.



Example of protocol





Numerical data : Nucleus

Recognition : Nucleus





NFkB is one of the famous transcription factor of DNA. NFkB plays a key role in regulating the immune response and inflammation and is attracting attention as a tumor therapy and anti-inflammatory drug target. NFkB is located in the cytoplasm with IkB which is inhibitory protein. Once the signaling pathway has been activated by the cytokine stimulation via cell membrane receptor, dissociate IkB from NFkB and activate NFkB. Then NFkB translocate into the nucleus to bind specific sequence of DNA, which induce inflammation. Nucleus and intracellular NFkB level indicates protein level between cytoplasm and nucleus.







Numerical data : Cellbody Numerical data : Dots (Volume, Intensity, Morphology, etc.) (Volume, Intensity, Morphology, etc.) (Volume, Intensity, Morphology, etc.)

Precise separation of localization by the confocal unit

Membrane translocation

Try time lapse imaging! New

Keep cells happier in incubator to see how they react on live. Low photo toxicity and photo bleaching offers time lapse imaging of live cell to see what they are.





Time lapse analysis : Apoptosis





Spread HeLa cells to 96well microplate with 10,000 cells/well.

Stain with Hoechst33342 (1 µg/ml, 30 min, 37 ℃) and treat with Staurosporine (0 - 10µM) and capture image every 15 min. Recognize DNA fragmentation area of nuclei at Staurosporine 10µM treatment.



Measurements of Volume, Intensity and Morphology

Cellcycle, Apoptosis

Example of protocol



Numerical data (Volume, Intensity, Morphology, etc.)

Numerical data (Volume, Intensity, Morphology, etc.)







Time lapse analysis of colony size and individual cells allow to monitor colony formation state. CQ1's image can perform image acquisition with low photo-toxicity. Data provided by Kyoji Horie, ph.D, Physiology II, Nara Medical University







Measurement (Time:10)

Numerical data (Volume, Intensity, Morphology, etc.)

Time lapse analysis : ESC colony

Time course measurement allows to monitor cell colony growth

Want to try the measurement again...

Cells can be imaged at culture plate, no need to prepare single-cell suspension, and you can use same sample to different measurement.

Image and analysis data are associated together and its help to pick up tiny difference.







Example of CTC quantitate (spiking experiment). CTC : CD45 is only Negative. Data provided by Yusuke Tomita, Min-Jung Lee, Jane B Trepel , Developmental Therapeutics Branch, National Cancer Institute, National Institutes of Health, , Bethesda, MD 20892 USA

CTCs are tumor cells which circulate in peripheral blood. Developed tumors metastasize through the bloodstream and lymph fluid. Therefore, tumor cells exist in the bloodstream when metastasis occurs. The detection of CTCs makes it possible to diagnose recurrence and metastasis at an early tumor stage. CTCs' numbers are very small as only less than 100 CTCs are contained in more than 1x10⁶ of blood cells in 10 ml of cancer patient's blood. Therefore it is difficult to detect CTCs with a flow cytometer because they detect CTCs as noise. However, it is very easy to detect rare CTCs with an Image cytometer.



2 (0.001%)

CTCs (count)

Template CTC

You can detect multiple marker expression of the cell. Not only for circulating tumor cells, but also for the other specific marker can be detected.

Example of protocol





Numerical data : Nucleus (Volume, Intensity, Morphology, etc.)





Cell cycle analysis in relation to H3Ser10P immunofluorescence by utilizing the CQ1' s multi-color channel capabilities. Histone molecules are phosphorylated during cell cycle progression with phosphorylation of the 10th serine of histone H3 being one of the well characterized events of late-G2 to M progression.



Template CellCycle

You can detect cell cycle to verify drug treatment efficiency. This is available by the flow cytometer, but CQ1 can analyze more items which typical at the image cytometer.







Cell cycle analysis: M-phase inhibitor