

High Sensitivity SSC and Fluorescence Detection Using the NovoCyte Advanteon Flow Cytometer

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Abstract

This Application Note describes the ability of the Agilent NovoCyte Advanteon flow cytometer to resolve small particles, including cell-derived extracellular vesicles (EVs) and beads. The NovoCyte Advanteon, equipped with advanced detectors, provides high sensitivity side-scatter (SSC) and fluorescence detection to give superior results to those of traditional flow cytometers.

Introduction

The importance of cell-derived EVs is recognized in multiple biological processes, and is a topic of increasing relevance in biomedical research. EVs are cell-derived membranous vesicles released from cells under normal and pathological conditions. This includes exosomes, microvesicles, and apoptotic bodies according to their cellular origin and size. Exosomes are 50 to 150 nm of endocytic origin. Microvesicles bud from the cell surface and range in size from 50 nm to 1 μm , and apoptotic bodies are 0.8 to 5 μm vesicles released from cells undergoing programmed cell death. EVs are important for intracellular communication, and have been found in blood, other biological fluids, and cultured cell supernatant. EVs have been associated with a wide range of conditions including cancer metastasis, blood disorders, cardiovascular disease, infectious disease, and others. The therapeutic potential of EVs as a cell-free treatment for cancer and other pathologies has been investigated extensively. Current EV detection methods use techniques such as western blotting and enzyme-linked immunosorbent assay (ELISA) to determine bulk protein concentration in purified particles, which prevents readout of individual samples. Including flow cytometric assays in small particle research could enhance current detection methods,

with the capability to analyze individual particles in significant numbers and measure surface protein levels. However, traditional flow cytometers have difficulty resolving EVs due to their small diameter and variable composition. Technological advancements in flow cytometry, as with the NovoCyte Advanteon, have increased the capacity to detect small particles, such as EVs. This investigation determines the ability of the NovoCyte Advanteon flow cytometer to resolve small particles using beads and cell-derived EVs.

Experimental

The NovoCyte Advanteon can detect small particle beads with sensitive SSC detection

To determine the capability of the NovoCyte Advanteon to resolve small particles, we first performed an analysis of three commercially available small particle bead mixes:

- Bang's laboratories NIST-traceable size standards
- ApogeeMix beads
- BioCytex-Plus FSC and SSC hybrid microspheres

These each consist of beads ranging in size from 100 to 1,300 nm. Equipped with an avalanche photodiode for SSC instead of the traditional photodiode detector, the NovoCyte Advanteon was able to resolve beads as small as 100 nm using SSC alone (Figure 1).

Sensitive resolution of small biological particles by flow cytometry

The refractive index (RI) of membrane vesicles is different than particle beads, therefore it is important to determine the capability of the NovoCyte Advanteon to detect biologically derived EVs. We analyzed purified lipoparticles derived from HEK293T green fluorescent protein (GFP) expressing cells, the release of which is induced by Murine leukemia virus (MLV). These lipoparticles are approximately 240 nm determined by dynamic light scattering (Figure 2A). Lipoparticles were separated from the background (Figure 2B) by SSC as well as GFP fluorescence. In addition, the NovoCyte Advanteon features a syringe pump precisely measuring sample volumes, allowing direct absolute counting of lipoparticles. Figure 2C shows the results of a serial dilution of MLV-induced lipoparticles. With small particle detection, it is important to ensure that single particles are analyzed. If particles are run at too high of a concentration, the particles can be too close together and multiple particles may be detected as a single event. This phenomenon is known as coincidence or the swarm effect. Individual particles that would normally register below the detection threshold are detected as a cumulative event, enabling them to reach the trigger threshold. This effect distorts the data, altering the detected sample concentration and phenotype. One way to control for this is to perform serial dilutions of the sample, to ensure single particle detection. If there is no coincidence, a linear decrease in sample concentration should be observed with serial dilutions. Based on the measured absolute event count, the actual dilution factor correlated well with the calculated dilution factor (Figure 2C), confirming that the events detected were not due to coincidence. These data demonstrate the capacity of the NovoCyte Advanteon to detect small membranous particles.

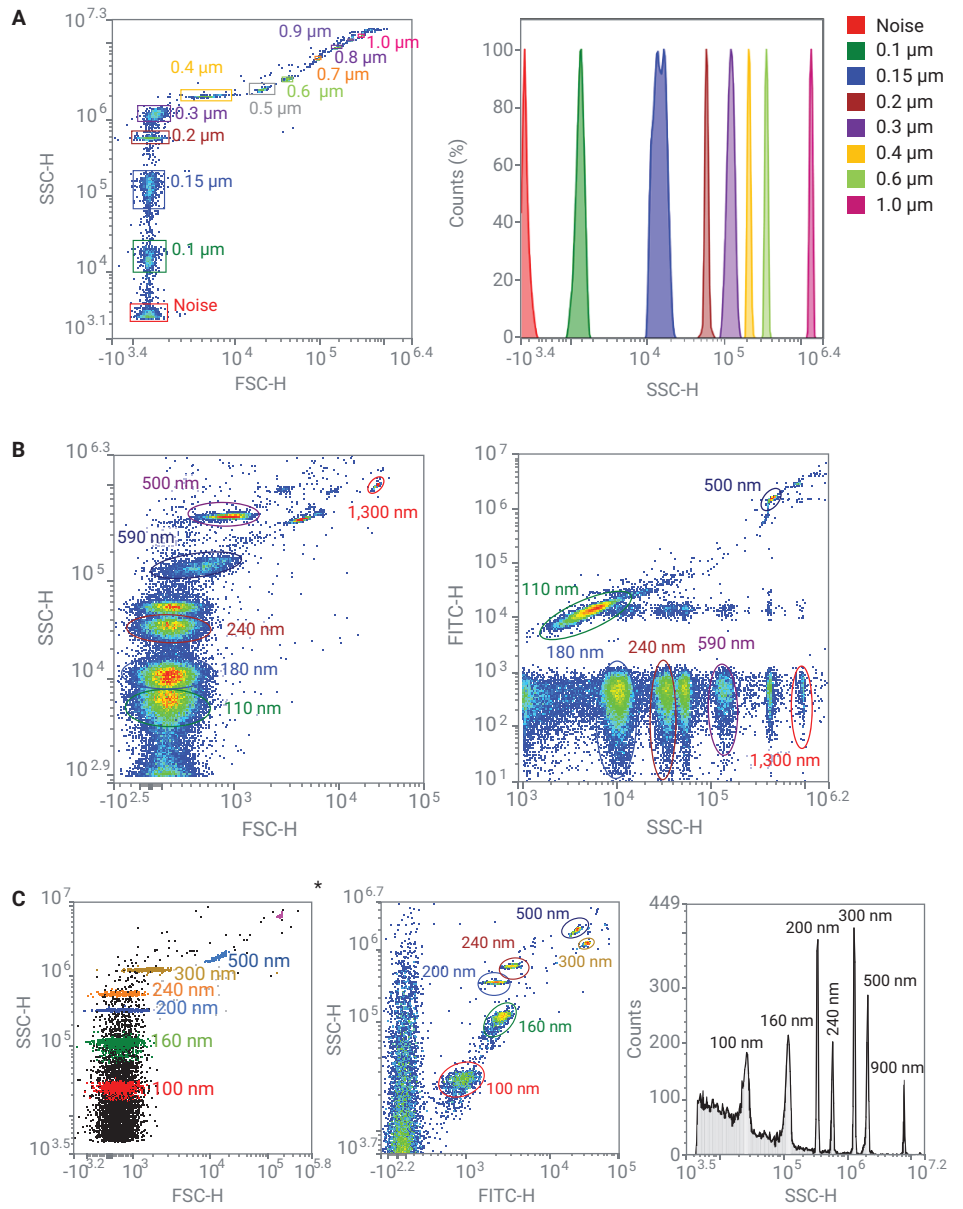


Figure 1. Small particle bead detection on the Agilent NovoCyte Advanteon. Three commercially available small particle beads were used to demonstrate the high sensitivity of the NovoCyte Advanteon side scatter detection (SSC): Bang's Laboratories NIST traceable size standards (A), ApogeeMix beads (B), and BioCytex Megamix-Plus (C). Particles as small as 100 nm were easily resolved from the background. Instrument settings: SSC-H gain 1,000, FSC-H gain 1,000, threshold SSC-H greater than 1,000.

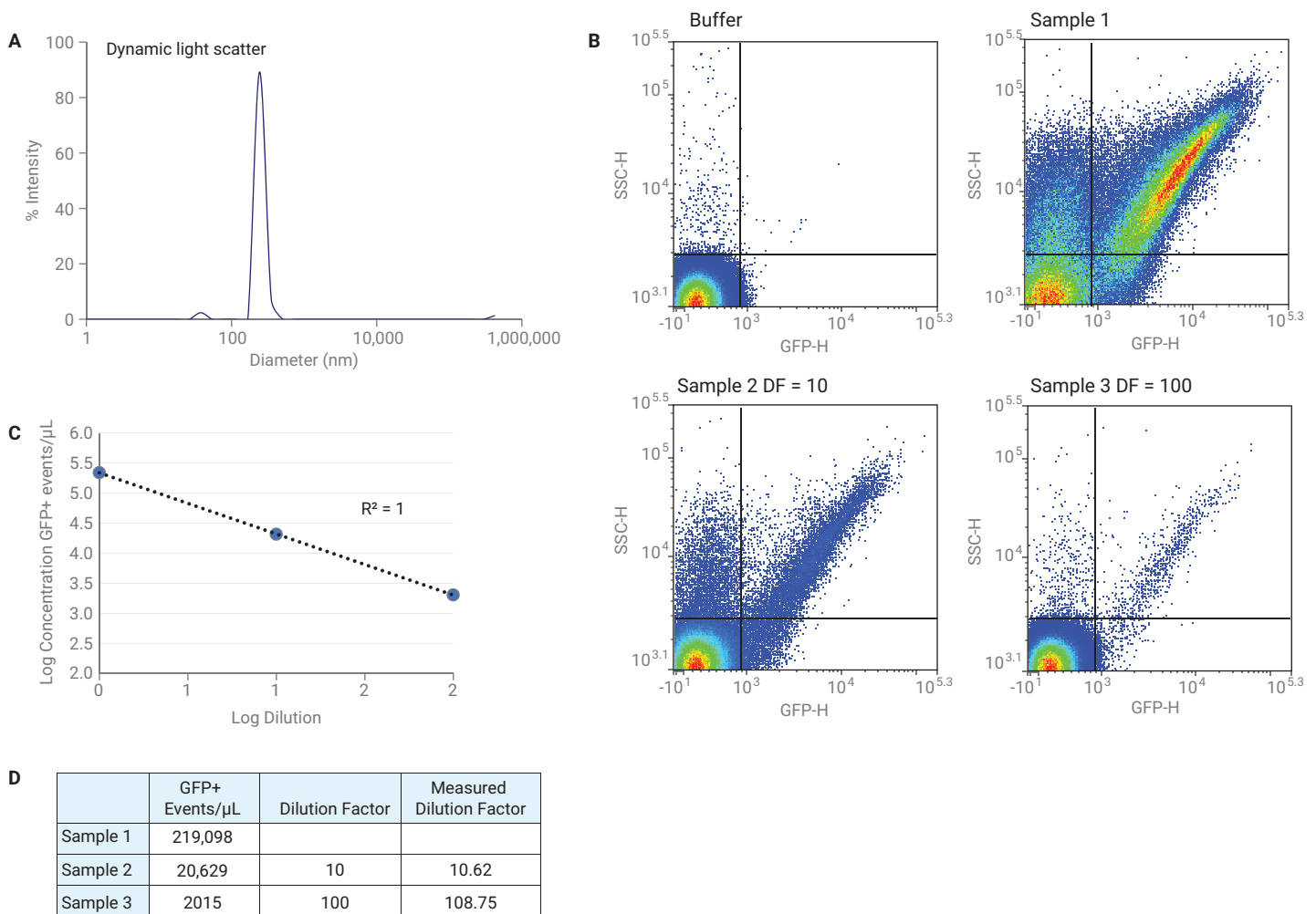


Figure 2. MLV-induced lipoparticle detection using the Agilent NovoCyte Advanteon. (A) Dynamic light scattering of lipoparticles to determine size. (B) SSC-H versus GFP-H plots of serially diluted lipoparticle fraction. Sample 1: 1:1,000, Sample 2: 1:10,000, Sample 3: 1:100,000. (C,D) Linearity of absolute count of GFP+ events obtained by volumetric counting of the sample dilution series, (C,D) linearity curve of dilution factor and concentration of GFP+ events/μL. Instrument settings: SSC-H gain 900, threshold FITC-H greater than 220, 14 μL/min.

Platelet activation increases released EV in plasma

Upon activation, platelets release procoagulant EVs, for example during thrombosis or inflammation. These particles range from 50 nm to 1 μm, however, most plasma-derived EVs are 100 to 250 nm.

To induce EV release, platelets were activated with ADP. After activation, total EVs were isolated through ultracentrifugation, and stained with anti-CD9, anti-CD61 antibodies, and Annexin V to label EVs (Figure 3). CD61 is a highly prevalent surface

receptor on platelets, and identifies platelet-originated EVs, while Annexin V and CD9 are commonly used as universal markers for EVs. EV concentration increased by more than five times after platelet activation with ADP (Figure 3H). The highest concentration of EVs was identified by expression of CD9 (Figures 3A and 3B). Most CD9+ EVs also expressed Annexin V and CD61, confirming that they are derived from platelets. A lower concentration of EVs was identified by surface expression of CD61 or Annexin V, demonstrating the heterogeneity of EVs present in the plasma. To confirm

that the particles detected were membrane vesicles and not protein aggregates or other nonmembranous structures, samples were incubated with a detergent, Triton-X, for one hour to disrupt the plasma membrane and eliminate any vesicles present in the sample. Following Triton-X treatment, previously detected EVs disappeared and only background events were present, confirming that the events detected were EVs. In summary, platelet-derived EVs are easily detected using the NovoCyte Advanteon by either platelet or EV-specific markers.

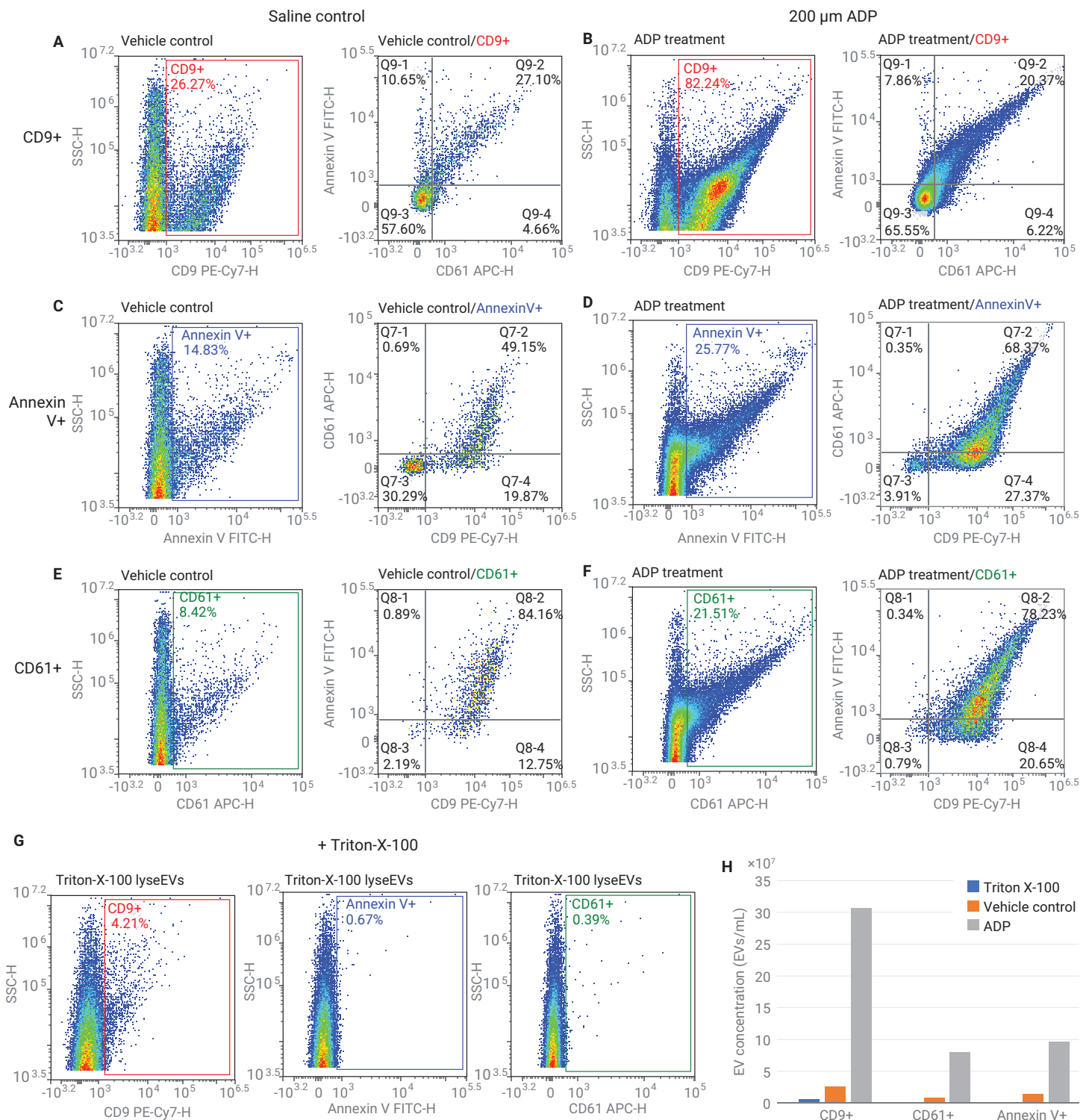


Figure 3. Increased EV release by platelet activation can be resolved by flow cytometry. Donor blood was collected using a sodium citrate blood collection tubes. Whole blood was treated either with 200 μ m ADP or saline alone for 30 minutes. EVs were then isolated by centrifugation: 2,500 \times g for 15 minutes twice, then the supernatant was centrifuged at 110,000 \times g for 90 minutes twice. This was followed by staining with α CD9 PE-Cy7, α CD61 APC antibodies, and Annexin V FITC. EVs were identified by CD9 (A,B) and Annexin V (C,D), and CD61 (E,F) in the saline-treated control (A,C,E) or ADP-treated samples (B, D,F). (G) Identified EVs were analyzed for co-expression of other EV markers (A-F, right plots). A portion of the vesicles was incubated with 1% Triton-X-100 on ice for one hour. (H) Graph with EV concentration under the conditions described in the text. Instrument settings: SSC-H gain 1,000, threshold SSC-H greater than 1,000.

Conclusion

EVs are released from various cells and found in biological fluids and cell culture supernatant. They are easy to obtain and carry important biological information.

They are potential biomarkers for cancer, infectious disease, and pathological conditions, indicating their value for disease research. The Agilent NovoCyte Advanteon flow cytometer, equipped with advanced detectors, provides high-sensitivity SSC and fluorescence detection, overcoming many of the obstacles of small particle detection in traditional flow cytometers.

References

1. Shao, H. *et al.* New Technologies for Analysis of Extracellular Vesicles. *Chem. Rev.* **2018**, *118*(4), 1917–1950.
2. Aatonen, M. T. *et al.* Isolation and Characterization of Platelet-Derived Extracellular Vesicles. *J. Extracell. Vesicles* **2014**, *3*.
3. Osteikoetxea, X. *et al.* Differential Detergent Sensitivity of Extracellular Vesicle Subpopulations. *Org. Biomol. Chem.* **2015**, *13*(38), 9775–9782.
4. Becker, A. *et al.* Extracellular Vesicles in Cancer: Cell-To-Cell Mediators of Metastasis. *Cancer Cell* **2016**, *30*(6), 836–848.
5. Xu, R. *et al.* Extracellular Vesicle Isolation and Characterization: Toward Clinical Application. *J. Clin. Invest.* **2016**, *126*(4), 1152–1162.
6. Aatonen, M. T. *et al.* Isolation and Characterization of Platelet-Derived Extracellular Vesicles. *J. Extracell. Vesicles* **2014**, *3*.

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